

Short paper

Open Access

Improved gene delivery to human saphenous vein cells and tissue using a peptide-modified adenoviral vector

Lorraine M Work¹, Paul N Reynolds² and Andrew H Baker*¹

Address: ¹BHF Glasgow Cardiovascular Research Centre, Division of Cardiovascular & Medical Sciences, University of Glasgow, 44 Church Street, Glasgow, G11 6NT, UK and ²Royal Adelaide Hospital Chest Clinic and Department of Medicine, University of Adelaide, Adelaide, South Australia, Australia

Email: Lorraine M Work - lmw3u@clinmed.gla.ac.uk; Paul N Reynolds - paul.reynolds@adelaide.edu.au; Andrew H Baker* - ab11f@clinmed.gla.ac.uk

* Corresponding author

Published: 08 October 2004

Received: 10 September 2004

Genetic Vaccines and Therapy 2004, **2**:14 doi:10.1186/1479-0556-2-14

Accepted: 08 October 2004

This article is available from: <http://www.gvt-journal.com/content/2/1/14>

© 2004 Work et al; licensee BioMed Central Ltd.

This is an open-access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

The establishment of efficient gene delivery to target human tissue is a major obstacle for transition of gene therapy from the pre-clinical phases to the clinic. The poor long-term patency rates for coronary artery bypass grafting (CABG) is a major clinical problem that lacks an effective and proven pharmacological intervention. Late vein graft failure occurs due to neointima formation and accelerated atherosclerosis. Since CABG allows a clinical window of opportunity to genetically modify vein *ex vivo* prior to grafting it represents an ideal opportunity to develop gene-based therapies. Adenoviral vectors have been frequently used for gene delivery to vein *ex vivo* and pre-clinical studies have shown effective blockade in neointima development by overexpression of candidate therapeutic genes. However, high titers of adenovirus are required to achieve sufficient gene delivery to provide therapeutic benefit. Improvement in the uptake of adenovirus into the vessel wall would therefore be of benefit. Here we determined the ability of an adenovirus serotype 5 vector genetically-engineered with the RGD-4C integrin targeting peptide inserted into the HI loop (Ad-RGD) to improve the transduction of human saphenous vein smooth muscle cells (HSVSMC), endothelial cells (HSVEC) and intact saphenous vein compared to a non-modified virus (Ad-CTL). We exposed each cell type to virus for 10, 30 or 60 mins and measured transgene at 24 h post infection. For both HSVSMC and HSVEC Ad-RGD mediated increased transduction, with the largest increases observed in HSVSMC. When the experiments were repeated with intact human saphenous vein (the ultimate clinical target for gene therapy), again Ad-RGD mediated higher levels of transduction, at all clinically relevant exposures times (10, 30 and 60 mins tissue:virus exposure). Our study demonstrates the ability of peptide-modified Ad vectors to improve transduction to human vein graft cells and tissue and has important implications for gene therapy for CABG.

Text

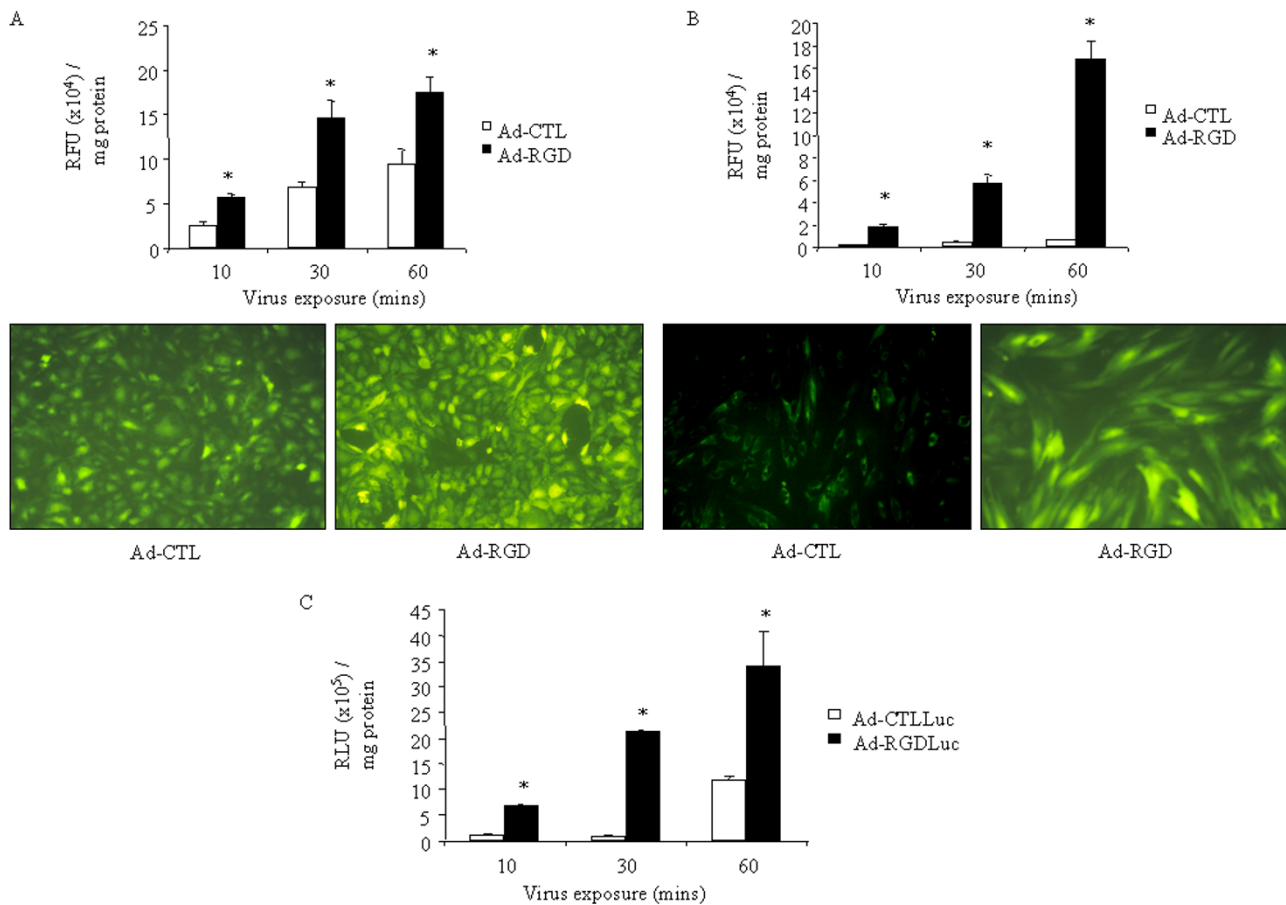
Long term patency rates for CABG using autologous saphenous vein are poor, showing 1, 5 and 10 years post-CABG rates of 93%, 74% and 41%, respectively [1] and

therefore represent a significant clinical problem. Long-term failures are due to neointima formation and superimposed atherosclerosis [2-4], a pathology that lacks a suitably efficient pharmacological therapy. Significant

contributions of vascular smooth muscle cell (SMC) proliferation and migration have been documented [2-4]. Anti-proliferative strategies are in phase III clinical trial using decoy oligonucleotides to the transcription factor E2F, a strategy that has shown considerable promise pre-clinically [5,6] and also in early stage human trials [7]. We, and others have adopted the alternate strategy of gene therapy to prevent CABG failure [8]. CABG is an "ideal" clinical scenario for gene therapy since saphenous vein can be genetically modified *ex vivo* following leg harvesting and prior to coronary grafting. This unique "clinical window" has clear safety advantages over *in vivo* gene delivery since excess vector can be removed from the graft prior to coronary grafting. However, the clinical window is short (likely 10–60 minutes) and therefore necessitates the use of an efficient vector system for gene delivery. Adenoviral vectors have proven efficient for gene delivery in this context [9] although high titers are required to provide sufficient levels of gene delivery to achieve therapeutic gain using transgenes such as tissue inhibitor of metalloproteinases-3 [9], endothelial nitric oxide synthase [10] and p53 [11]. The latter study defined the rationale behind use of adenoviral vectors since long-term benefit on graft remodelling was shown at 3 months, even though the virus was only present for 2–4 weeks post grafting [11]. Any improvement in gene delivery above that mediated by adenoviral serotype 5 vectors would be very encouraging for clinical translation of pre-clinical therapies. To this end, a number of strategies have emerged including fiber switching (pseudotyping) and modification of adenovirus type 5 fibers with targeting peptides. Pseudotyping the fiber from adenovirus serotype 16, which binds CD46 [12], dramatically improves transduction to vascular cells including intact human saphenous vein allowing lower doses of vector to be used to achieve attractive levels of gene delivery to grafts *ex vivo* [13]. Likewise, gene delivery to vascular smooth muscle cells can be enhanced by incorporation of cell targeting peptides isolated by phage display into the HI loop of the adenovirus fiber [14], the preferred site for peptide insertion [15]. In the context of improved gene delivery mediated by the RGD-4C peptide, which was isolated by phage display and targets α_v integrins [16], this has been shown for rabbit grafts [17] although the vast majority of data is based on gene delivery for cancer [18]. Since SMC show poor coxsackie and adenovirus receptor (CAR) availability [19], it is particularly relevant that the RGD-4C peptide may circumvent CAR deficiency on target cells to improve levels of transduction. In this study, we assess the ability of RGD-4C-modified adenovirus serotype 5 vectors to enhance gene delivery to human saphenous vein SMC and EC as well as to intact human saphenous vein *ex vivo*, the ultimate clinical target.

HSVEC were obtained by enzymatic collagenase digestion of human saphenous vein and maintained in endothelial cell complete media (TCS CellWorks, UK) supplemented with 20% (v/v) foetal calf serum (FCS; PAA laboratories, UK). HSVSMC were grown from medial explants from the same material and maintained in Dulbecco's modified Eagle's medium (DMEM) with 4500 mg/l glucose supplemented with 20% (v/v) FCS and 100 IU/ml penicillin, 100 μ g/ml streptomycin and 2 mmol/l L-Glutamine. All cells were grown in a humidified atmosphere with 5% CO₂ at 37°C. Cells were plated to reach 80% confluence 24 hours later. HSVEC or HSVSMC were infected in 96 well plates with increasing doses [plaque forming units (pfu) / cell] of Ad vectors for 10, 30, 60 mins at 37°C. The cells were washed twice in PBS and the media changed. 24 hours post-infection, the cells were again washed in PBS, lysed in PBS/0.2% Triton-X-100 and transduction quantified using the Wallac 1420 (Victor2) Multilabel Counter with recombinant eGFP (Clontech, Basingstoke, UK) as a standard. Reporter gene expression was normalised for total protein using the bicinchoninic acid (BCA) protein assay (Perbio, UK) with bovine serum albumin as standard, measured using a VICTOR2 plate reader. Exposure of HSVEC to Ad-CTL or Ad-RGD [50 plaque forming units (pfu) / cell] resulted in a time-dependent increase in the level of transduction (Figure 1A). At each time point studied (10, 30 or 60 mins), Ad-RGD mediated a significantly enhanced level of transgene expression compared to Ad-CTL (Figure 1A). Fluorescent microscopy demonstrated that control levels of infection with Ad-CTL were relatively high in HSVEC but further enhanced using the RGD-modified Ad (Figure 1A) at all time points tested. This is consistent with HSVEC expressing moderate CAR levels [14,19] allowing transduction of cells by Ad-CTL but the RGD-4C vector can further improve virus uptake. In HSVSMC, Ad-RGD again mediated a marked and significant enhancement in levels of transgene expression at all time points studied (Figure 1B). HSVSMC were much less permissive to non-modified Ad-CTL infection (Figure 1B), consistent with our previous observations [14], but enhanced with Ad-RGD to near 100% transduction in HSVSMC by fluorescence microscopy (Figure 1B). Again, this effect was evident at all virus:cell exposure time points – 10, 30 and 60 mins. For both HSVEC and SMC similar RGD-4C-mediated increases were observed with different viral doses (10 and 100 pfu/cell; not shown) thereby showing both time- and dose-dependence.

Based on the above we therefore assessed transduction in intact human saphenous vein. In order to quantify transgene expression accurately in tissue extracts we used luciferase-expressing viruses. Intact human saphenous veins were cleaned of surrounding connective tissue and cut into rings 3–4 mm in length. During preparation and infection, veins were maintained in wash medium (RPMI

**Figure 1**

Transduction of saphenous vein cells and intact tissue. Ad-CTL and Ad-RGD expressing eGFP were incubated with (A) HSVEC and (B) HSVMC for different times and gene expression quantified and normalised to protein. Representative fluorescent images are shown. (C) Intact human saphenous vein was incubated with luciferase-expressing vectors and expression quantified. *Indicates $p < 0.05$ vs Ad-CTL.

supplemented with 100 IU/ml penicillin, 100 μ g/ml streptomycin and 2 mmol/l L-glutamine). Individual vein rings were incubated with Ad vectors for 10, 30 or 60 minutes (1×10^9 pfu / ring) before being washed twice in PBS and maintained in organ culture for 5 days. Rings were maintained in wash medium supplemented with 30% (v/v) FCS. Vein rings were snap frozen in liquid nitrogen and homogenised using a mortar and pestle for determination of reporter gene expression 5 days post-infection. *Ex vivo* homogenates were suspended in 100 μ L reporter lysis buffer (RLB) and kept on ice for 1 hour before supernatants were analysed for luciferase expression using the Luciferase Assay System (Promega). 96 well plates were prepared using 10 μ L/well of homogenate suspension diluted to a total volume of 100 μ L with RLB. 100 μ L of Luciferase Assay Reagent was added to each well and the

plate immediately read for 10 seconds per well. Detection was achieved using a Wallac 1420 (VICTOR2) Multilabel Counter with recombinant luciferase (Promega) as a standard and normalised for total protein. Ad-RGDLuc mediated a time-dependent increase in the level of transgene expression that was evident at all exposure times studied – 10, 30 and 60 minutes (Figure 1C). This demonstrates that the RGD-4C-modification of Ad vectors can increase transduction to human saphenous vein, especially at short exposure times. The kinetics of virus binding in relation to time is therefore improved through the RGD-4C peptide and has direct implications for the design of gene therapy vectors for use in human CABG gene therapy procedures in the future. Although we have previously shown that non-modified Ad vectors transduce both endothelial and smooth muscle cells during graft

gene delivery [9], and here show increased transduction of both cell types *in vitro* with RGD-modification, it will be important to fully define the uptake of the RGD-modified virus in the intact vein at the cellular level by immunotechniques. In broader terms, the design and tailoring of viruses for individual cardiovascular gene therapy applications is an important aspect of translation from pre-clinical to clinical gene therapy.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

LMW performed all isolated cell culture and vein transduction experiments. PNR produced the viruses and AHB supervised all work as principle investigator

The authors thank Nicola Britton and Margaret Cunningham for technical assistance. This work was supported by the Biotechnology & Biological Sciences Research Council (E17190 to A.H.B) and the British Heart Foundation (PG03/031 to A.H.B.)

References

- Eagle KA, Guyton RA, Davidoff R, Ewy GA, Fonger S, Gardner TJ, Gott JP, Herrmann HC, Marlow RA, Nugent WC, O'Connor GT, Orszulak TA, Rieselbach RE, Winters WL, Yusuf S, Gibbons RJ, Alpert JS, Eagle KA, Gardner TJ, Garson A, Gregoratos G, Russell RO, SC Smith, McEntee CW, Elma MA, Pigmann GC, Starke RD, Taubert KA: **ACC/AHA guidelines for coronary artery bypass graft surgery: A report of the American College of Cardiology/American Heart Association task force on practice guidelines (committee to revise the 1991 guidelines on coronary artery bypass graft surgery).** *J Am Coll Cardiol* 1999, **34**:1262-1342.
- Cox JL, Chiasson DA, Gotlieb AI: **Stranger in a strange land: the pathogenesis of saphenous vein graft stenosis with emphasis on structural and functional differences between veins and arteries.** *Prog Cardiovasc Dis* 1991, **34**:45-68.
- Bryan AJ, Angelini GD: **The biology of saphenous vein graft occlusion: etiology and strategies for prevention.** *Curr Opin Cardiol* 1994, **9**:641-649.
- Davies MG, Hagen P-O: **Pathobiology of intimal hyperplasia: a review.** *Eur J Vasc Surg* 1995, **9**:7-18.
- Mann MJ, Gibbons GH, Kernoff RS, Diet FP, Tsao PS, Cooke JP, Kaneda Y, Dzau VJ: **Genetic engineering of vein grafts resistant to atherosclerosis.** *Proc Natl Acad Sci USA* 1995, **92**:4502-4506.
- Mann MJ, Gibbons GH, Tsao PS, vonderLeyen HE, Cooke JP, Buitrago R, Kernoff R, Dzau VJ: **Cell cycle inhibition preserves endothelial function in genetically engineered rabbit vein grafts.** *J Clin Invest* 1997, **99**:1295-1301.
- Mann MJ, Whittemore AD, Donaldson MC, Belkin M, Conte MS, Polak JF, Orav EJ, Ehsan A, Dell'Acqua G, Dzau VJ: **Ex-vivo gene therapy of human vascular bypass grafts with E2F decoy: the PREVENT single-centre, randomised, controlled trial.** *The Lancet* 1999, **354**:1493-1498.
- Baker AH, Mehta D, George SJ, Angelini GD: **Prevention of vein graft failure: potential applications for gene therapy.** *Cardiovasc Res* 1997, **35**:442-4450.
- George SJ, Lloyd CT, Angelini GD, Newby AC, AH Baker: **Inhibition of late vein graft neointima formation in human and porcine models by adenovirus-mediated overexpression of tissue inhibitor of metalloproteinase-3.** *Circulation* 2000, **101**:296-2304.
- West Nick E.J., Qian HuSheng, Guzik Tomasz J., Black Edward, Cai Shijie, George Samuel E., Channon Keith M.: **Nitric oxide synthase (nNOS) gene transfer modifies venous bypass graft remodeling: effects on vascular smooth muscle cell differentiation and superoxide production.** *Circulation* 2001, **104**:1526-1532.
- Wan S, George SJ, Nicklin SA, Yim APC, Baker AH: **Overexpression of p53 increases lumen size and blocks neointima formation in porcine interposition vein grafts.** *Mol Ther* 2004, **9**:689-698.
- Gaggar A, Shayakhmetov DM, Lieber A: **CD46 is a cellular receptor for group B adenoviruses.** *Nat Med* 2003, **9**:1-15.
- Havenga MJE, Lemckert AAC, Grimbergen JM, Vogels R, Huisman LGM, Valerio D, Bout A, Quax PHA: **Improved adenovirus vectors for infection of cardiovascular tissues.** *J Virol* 2001, **75**:3335-3342.
- Work LM, Nicklin SA, Brain NJR, Dishart KL, Von Seggern DJ, Hallek M, Buning H, Baker AH: **Development of efficient viral vectors selective for vascular smooth muscle cells.** *Mol Ther* 2004, **9**:198-208.
- Dmitriev I, Krasnykh V, Miller CR, Wang MH, Kashentseva E, Mikheeva G, Belousova N, Curiel DT: **An adenovirus vector with genetically modified fibers demonstrates expanded tropism via utilization of a coxsackievirus and adenovirus receptor-independent cell entry mechanism.** *J Virol* 1998, **72**:9706-9713.
- Pasqualini R, Koivunen E, Ruoslahti E: **A peptide isolated from phage display libraries is a structural and functional mimic of an RGD-binding site on integrins.** *J Cell Biol* 1995, **130**:1189-1196.
- Hay CM, De Leon H, Jafari JD, Jakubczak JL, Mech CA, Hallenbeck PL, Powell SK, Liao G, Stevenson SC: **Enhanced gene transfer to rabbit jugular veins by an adenovirus containing a cyclic RGD motif in the HI loop of the fiber knob.** *Journal of Vascular Research* 2001, **38**:315-323.
- Nicklin SA, Baker AH: **Tropism-modified adenoviral and adeno-associated viral vectors for gene therapy.** *Curr gene ther* 2002, **2**:273-2293.
- Nicklin SA, Von Seggern DJ, Work LM, Pek DCK, Dominiczak AF, Nemerow GR, Baker AH: **Ablating adenovirus type 5 fiber-CAR binding and HI loop insertion of the SIGYPLP peptide generate an endothelial cell-selective adenovirus.** *Mol Ther* 2001, **4**:534-5542.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

